

## STRP™ HIV-1 Detection Kit (Short Protocol)

Cat. No.: PR8249C  
 Shipment: Wet Ice

Storage: -20°C  
 Quantity: 20 Reactions

This kit is destined for qualitative detection of HIV-1 RNA in the serum and plasma of Human blood by the method of Single tube RT reaction, followed by nested PCR. CinnaGen HIV-1, RT-PCR detection kit may be used in clinical medicine to detect HIV-1 RNA.

### Kit Contents

1. RNXTM-Plus	9 ml	5. RT Enzyme	20 µl
2. Mix I	780 µl	6. Taq DNA Poly.	10 µl
3. Mix II	440 µl	7. DNA Pos.	25 µl
4. DEPC-Water	600 µl	8. Mineral oil	2 ml

The Reagents Needed :

1. Chloroform
2. Isopropanol
3. 70% Ethanol

### A) RNA Extraction

Perform in Pre-Amplification 1, Specimen & Control Area.

1. Add 50 µl Serum or Plasma to 450 µl cold RNXTM-Plus solution. Vortex the sample to dissolve the clamps. Incubate for 10 min on ice.
2. Add 100 µl of Chloroform, vortex (3-5 sec.) and centrifuge at 12000 g for 5 min.
3. Transfer the upper phase to new tube and add equal volume of Isopropanol (250-300 µl). Invert the tube 10 times and then incubate at -20°C for at least 20 min.
4. Centrifuge at 12000 g for 15 min.
5. Discard aqueous phase and add to the pellet 200 µl 70% Ethanol and invert 10 times, centrifuge at 12000 g for 5 min.
6. Discard aqueous phase and incompletely dry the pellet (RNA) for 20-30 min. at room temperature.
7. Dissolve RNA in 30 µl DEPC treated water, then follow the cDNA synthesis protocol within 3 hours of specimen preparation or store the processed specimens frozen at -70°C or colder for up to one month with no more than one freeze - thaw.

### B) Single tube cDNA Synthesis and first PCR Round

Perform in Pre-Amplification 2, Reagent Preparation Area.

Label PCR tubes for cDNA synthesis & first PCR, for test(s), positive and negative control.

1. Add the following reagents for each tube on ice (Mix & spin before use):

1X PCR Mix I	39 µl
RT Enzyme	1 µl
Taq DNA Polymerase	0.3 µl
Mineral oil	40 µl

2. Mix the mixture thoroughly by shaking and spin.
3. Close reaction tubes or place tray and reaction tubes in a resealable plastic bag and seal the bag securely, do not close reaction tubes at this time. Transfer tubes to Pre-Amplification 1 Area.
4. Place RNA tube at 95°C, 1 min. and then place on ice.
5. Add 5 µl RNA to each patient tube and Positive control to pos. tube and DEPC-Water to neg. tube.  
(The final volume of each reaction will be 45 µl)
6. Close tubes, spin the mixture on microfuge 3-5 sec. and transfer the tubes to preheated thermocycler and start the program:

### Cycling parameters:

42°C - 20 min.	93°C - 40 Sec.
94°C - 2 min.	Then 62°C - 40 Sec.
62°C - 40 sec.	72°C - 40 Sec.
72°C - 40 sec.	

### 1 cycle

### 20 cycles

Cycling parameters may need to be setup with some Thermocyclers. If so, please contact CinnaGen's Molecular Biology department.

### C) Second PCR Round

In Pre-Amplification 2, Reagent Preparation Area:

1. Add the following to PCR new reaction tube:

1X PCR Mix II	22 µl
Taq DNA Polymerase	0.2 µl
Mineral oil	20 µl

2. Close reaction tubes or place tray and reaction tubes in a resealable plastic bag and seal the bag securely, do not close reaction tubes at this time. Transfer tubes to Pre-Amplification 1 Area.
3. Add PCR product from first round 3 µl.  
(The final volume of each reaction tube will be 25 µl)
4. Transfer the tubes to preheated thermocycler and start the program:

Cycling parameters:

93°C - 40 sec
62°C - 40 sec
72°C - 40 sec
35 cycles

### D) Result Analysis

Performed in Post-Amplification Area.

Analyze amplified fragments by loading of 10 µl PCR product in 2% agarose gel directly without adding loading buffer. The presence of 174 bp fragments indicates positive test. In smear result with out specific fragment (174 bp), repeat the step B,C&D with 1/10 dilution of RNA (eg. 10 µl of RNA in 100 µl of DEPC-water).