



SinaClon First Strand cDNA Synthesis Kit

For Research Use Only

Cat. No.: RT0201

Store at: -20°C

Quantity: 50 Reactions

Shipment: Wet or Dry ice

Description

SinaClon First Strand cDNA synthesis Kit is specially designed to provide reliable synthesis of full-length cDNA from mRNA or total RNA templates. M-MuLV RNase-H synthesizes complementary DNA strand initiating from a specific primer, oligo(dT)₁₈ or random hexamer. The absence of RNase-H enhances the synthesis of long cDNA as the RNA strand does not degrade in DNA-RNA hybrid during first strand cDNA synthesis. The RNase inhibitor, supplied with the kit, effectively protects RNA from degradation at temperatures up to 60°C.

Kit Components(50 reactions):

M-MuLV Reverse Transcriptase (200 u/μl)	5000 u	
RNase Inhibitor (10 u/μl)	1000 u	
5X Buffer M-MuLV		200 μl
DTT (10 mM)	50 μl	
10 mM dNTPs mix		100 μl
Oligod(T) ₁₈	50 μl	
Random hexamer	50 μl	
DEPC-treated water	1 ml	

Storage and Stability

Stable at -20°C for 1 year if properly stored.

M-MuLV Storage Buffer:

20 mM Tris-HCl (pH 7.5), 200 mM NaCl, 10 mM EDTA, 1 mM DTT, Stabilizer, 5% (v/v) glycerol.

5X Buffer M-MuLV:

200 mM Tris-HCl (pH 8.3 at 25°C), 370 mM KCl, 10 mM MgCl₂.

RNase inhibitor Storage Buffer:

20 mM HEPES-KOH (pH 7.5), 50 mM KCl, 5 mM DTT, and 5% glycerol.

M-MuLV, Unit Definition:

One unit of the enzyme incorporates 1 nmol of dTTP into an acid-insoluble material in 10 minutes at 37°C using poly(rA). Oligo(dT)₁₈.

RNase inhibitor, Unit Definition:

1 u is defined as the amount of Ribonuclease inhibitor that inhibits the activity of 50 ng Ribonuclease A by 50%.

Preliminary Considerations:

Primers

Gene-specific primers, oligo(dT)₁₈ and random hexamer can be used in reverse transcription. Gene-specific primers transcribe only specific gene sequences and therefore increases the specificity. Gene-specific primers are used to synthesize specific cDNA from a pool of total RNA or mRNA and must be obtained by the user. Oligo (dT)₁₈ transcribes all poly (A) + mRNA, includes eukaryotic mRNA and viruses with poly (A) tail. Random hexamer initiate cDNA synthesis from the total RNA population (rRNA and mRNA). Therefore, using random



primers for first strand synthesis results in a greater complexity of the generated cDNA compared with the oligo(dT)₁₈ primer. As a consequence, the sensitivity and specificity of subsequent PCR reactions may be reduced. However, there are several applications where it is beneficial to use random primers, such as cDNA synthesis using mRNAs without a poly(A) tail, or cDNA synthesis using poly(A)-enriched RNA samples.

RNA templates

Quality and quantity of RNA templates determine the efficiency of reverse transcription process. The presence of minute amount of RNases can degrade the RNA and affect the cDNA length transcribed. To prevent RNases contamination, RNA purification has to be carried out in an RNase-free environment. Glassware, plasticware and reagents should be essentially RNase-free.

Recommended protocol for first strand cDNA synthesis

1. After thawing, mix and briefly centrifuge the components of the kit, Store on ice.
2. Prepare the RNA-primer mixture as below in a sterile, nuclease-free tube on ice.

component	Volume/ Concentration
Template: total RNA or poly A(+) mRNA	1 – 20 µg 0.1 – 2 µg
Primer: oligo(dT) ₁₈ or random Hexamers or gene-specific primer	1 µl 1 µl Volume depends on the primer stock concentration (10-20 pmol)
DEPC-treated water	Top up to 10 µl

3. Incubate the mixture at 37°C for 5 minutes and chill on ice for 2 minutes.
4. Briefly spin down the mixture.
5. Prepare the following cDNA Synthesis Mix in the order indicated:

Component	Volume
5X Buffer M-MuLV	5 µl
DTT	1 µl
M-MuLV Reverse Transcriptase	0.5 µl
RNase inhibitor	0.5 µl (20 u)
10 mM dNTP Mix	2 µl
DEPC-treated water	Top up to 10 µl

6. Add 10 µl of the cDNA Synthesis Mix into each RNA-primer mixture. Mix gently and centrifuge briefly.
7. Incubate at 42°C for 10 min.

Note: If random hexamer is used, incubation at 25°C for 10 min is needed prior to incubation at 42°C to prevent to dissociation of primer from template while the temperature increases. If gene specific primer or oligo (dT)₁₈ is used, this step can be ignored.

8. Terminate the reaction by incubate the tubes at 70°C for 5 min. Chill the tubes on ice and collect the solution by centrifuge the tube briefly.

9. The synthesized cDNA can be directly used in PCR, by addition of 1-2 µl of the cDNA reaction mixture to a 20 µl PCR reaction.

Quality Control

RT-PCR using 100 ng of control GAPDH RNA and GAPDH control primers generated a 497bp product visible on agarose gel after DNA Safe Stain staining.

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