

Helicobacter pylori PCR Detection Kit

Cat. No.: PR7843C

Storage: -20°C

Shipment: Wet Ice

Quantity: 20 Reactions

Description:

Helicobacter pylori PCR Detection kit is destined for qualitative detection of *Helicobacter pylori* DNA in culture or human gastric biopsy by the method of Polymerase Chain Reaction.

Kit Contents

The kit for 20 amplification reactions consists of:

1. 1X PCR MIX	400µl
2. Taq DNA polymerase (5 u/µl)	4µl
3. Mineral Oil	2.0ml
4. Positive Control	25µl
5. DNase Free Deionized Steril Water	5.0ml

- The Kit should be stored at - 20°C.

(Cycling parameters may need to be setup with some Thermocyclers. If so, please contact *CinnaGerts* molecular biology department.

Result Analysis

Performed in Post-Amplification Area

Analyze 10µl of amplified samples directly in a 1.5% agarose gel without adding loading buffer. The presence of 492 bp fragments indicates positive test.

For gel electrophoresis use of 100bp Ladder (Cat. No.: PR901644) is recommended.

DNA Extraction

Performed in Pre-amplification 1, Specimen & Control Area

Sample DNA can be extracted using CinnaGen DNP™ (Cat.No.: DN8115C) or CinnaPure™ (Cat.No.: PR881613).

PCR Protocol

Performed in Pre-Amplification 2, Reagent Preparation Area

- Take out the kit and unfreeze the tubes, then put all the tubes on ice. The final volume of each reaction will be 25µl.
- Label new 0.5 ml tubes for amplification reaction(s) for test(s), positive and negative control.
- Add the following reagents for each tube on ice:

1X PCR MIX	20µl
Taq DNA polymerase	0.2µl
- Mix the mixture thoroughly by shaking and spin.
- To each tube add one drop (20-25µl) mineral oil. Cap the reactions tube or Place the tube tray in a resealable plastic bag and seal the bag securely, don't cap tubes at this time. Next steps should be done at:

Pre -Amplification 1, Specimen & Control Preparation Area

- Add 5µl DNA* (Use specified pipette for sampling of DNA).
- Close tubes, spin the mixtures on microfuge for 3-5 seconds.
- Transfer the tubes to preheated thermocycler and start the program:

Cycling parameters:

First	Then
94°C - 45 sec	94°C - 20 sec
50°C - 20 sec	50°C - 20 sec
72°C - 30 sec	72°C - 30 sec
5 cycles	30 cycles