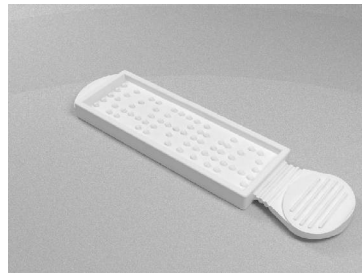


# SFM Dipslides

## Introduction:

Microbial test dipslide kit is an easy to use testing kit for the presence and count of microorganisms in liquids (such as urine, sputum, semen or wastewater...), different surfaces, cream, lotion and tissue slices of fruit and other plants and crops. The Health and Safety Executive's (HSE) recommends the use of dipslides to monitor the general activity of aerobic bacteria.



## Benefits:

- Simple application and use.
- Fast, easy and safe
- Precise and reliable
- Economical to use
- Can be used on liquids, surfaces and semi-solid materials
- Separate evaluation of bacteria, yeasts and molds

## Components and structure:

The dipslide test consists of two sterile culture medium on two-way plastic carrier .



## Diversity medium according to the customer such as:

Side 1:MacConkey agar

Side 2:Blood agar

Side 1:EMB agar

Side 2:Blood agar

Side 1:Nutrient agar

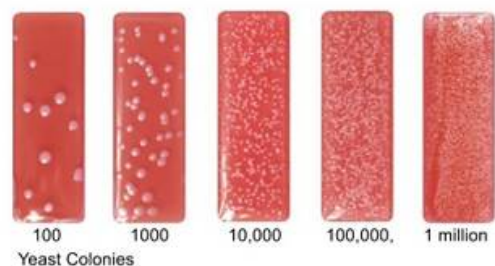
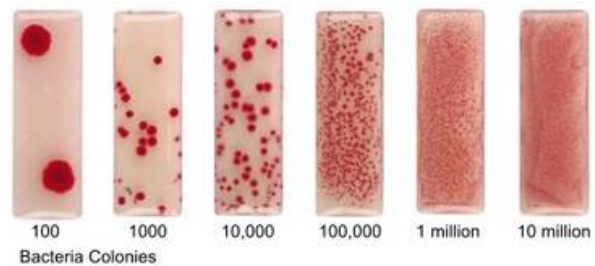
Side 2:Blood agar

Side 1:Nutrient agar

Side 2:EMB agar

## Contents:

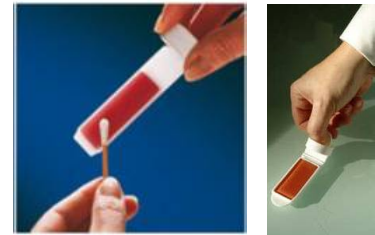
Each box contains 25 dipslides (in 50mL Conical Centrifuge sterile Tubes) and an user guide



## Application method:

### SURFACE Sampling Protocol

1. Remove the paddle from the vial. Do not touch the agar surfaces. Use aseptic techniques.
2. Firmly press the paddles (2X contact) against the test surface for a minimum of 3-5 seconds (15 seconds, optimal) for a 1:1 contact transfer.
3. Replace paddle in vial.
4. Incubate 25-35°C ± 2°C for 18-24



### LIQUID Sampling Protocol

#### DIRECT IMMERSION PROTOCOL – low viscous liquids

1. Mix liquid test sample.
2. Remove the paddle from the vial. Follow aseptic technique; do not touch the agar surfaces.
3. When taking the sample:
  - a. Pour 40mL of the sample into the vial (to the printed horizontal fill line). Dip the paddle into the 40mL volume liquid in the vial. Maintain a contact time of at least 15 seconds (30 seconds optimal). Both agar surfaces must be completely contacted.
  - b. Or dip the paddle into the sample directly. Maintain a contact time of at least 15 seconds (30 seconds optimal). Both agar surfaces must be completely contacted.
4. Allow excess fluid to drain off both paddle agar surfaces.
5. Replace paddle in vial.
6. Incubate 25-35°C ± 2°C for 18-24 hours.


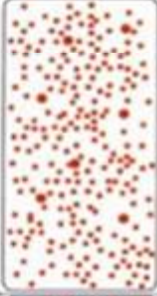

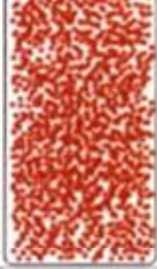



#### SPREAD Protocol – high viscous liquids or precise inoculation volumes

1. Mix liquid test sample.
2. Using aseptic technique, remove paddle from vial. Do not touch the agar surfaces.
3. Holding the contact agar surface on a horizontal plane, pipet 330µL (0.33mL) (deposit volume as a single drop (X)) approximately 1cm from the handle boundary
4. Position a sterile glass rod on the “handle” side of the drop (x) and bring it into contact with the drop creating a meniscus. Drag the glass tube over the paddle agar surface.
5. Replace paddle in vial.
6. Incubate 25-35°C ± 2°C for 18-24 hours



The evaluation chart shows colonies formed on the slides which correspond to different degrees of microbial contamination. The figures are shown in colony forming units per milileter (cfu/ml) on the included chart.

Total Colony Count (TCC)	Enumeration Panel Pictogram	Surface	Liquid
0		<1 cfu/cm <sup>2</sup>	<100 cfu/mL
1-5		<1 cfu/cm <sup>2</sup>	100 cfu/mL
10-50		1 cfu/cm <sup>2</sup>	10 <sup>3</sup> cfu/mL
100-500		10 cfu/cm <sup>2</sup>	10 <sup>4</sup> cfu/mL
>500		45 cfu/cm <sup>2</sup>	10 <sup>5</sup> cfu/mL
>1,000 (partial confluency)		80 cfu/cm <sup>2</sup>	10 <sup>6</sup> cfu/mL
>10,000		>100 cfu/cm <sup>2</sup>	10 <sup>7</sup> cfu/mL

**Storage and stability:**

Dip slide should be stored at about +4°C and protected from draught and the slides must not be allowed to freeze.

shelf life is 9-10 months; typically there is about 7-8 months shelf life from the time of manufacture.

**Warranty:**

**Sina Mehr Fajr Mazandaran Co.  
Sari.Iran**